Case Series

A Rare Extramedullary and Extralymphoid Presentation of Mixed Phenotypic Blastic Hematolymphoid Neoplasm: A Study of Two Cases

Abstract

Mixed phenotype acute leukemia (MPAL) is a rare hematolymphoid neoplasm, representing only 3%–5% of acute leukemia. Although MPAL has been sufficiently described in the literature, its extramedullary presentation as a solitary lesion without leukemic (bone marrow [BM]) involvement is rarely described. We are presenting two cases of mixed phenotypic blastic hematolymphoid neoplasms without leukemic involvement at disease presentation in 8-year-old female and 21-year-old male patients. Both the cases had extralymphatic bone involvement in the form of solitary bone lesion. Initially, there was no leukemic involvement in both the cases, but the second case progressed to acute leukemia during the course of the disease. On immunophenotypic evaluation, both the cases were difficult to categorize either into MPAL as the BM was not involved or into lymphoblastic lymphoma due to coexpression of myeloid differentiation. Therefore, we chose to classify them as a bi/mixed phenotypic blastic hematolymphoid neoplasm. Detailed immunophenotypic analysis either by immunohistochemistry or flow cytometric immunophenotyping is important for the diagnosis of such cases as they have a poor prognosis.

Keywords: Flow cytometric immunophenotyping, immunohistochemistry, mixed phenotype acute leukemia

Background

Mixed phenotype acute leukemia (MPAL) is a rare disease, representing only 3%-5% of acute leukemia of all age groups.^[1] MPAL comprises either single blasts population with the presence of more than one lineage-specific markers (formerly called as biphenotypic leukemia) or a presence of two distinct blast populations with two distinct lineage-specific markers (formerly called as bilineage leukemia).^[1] The World Health Organisation (WHO) definition of MPAL is based on the expression of lineage-specific markers of two or more lineages that surface/cytoplasmic include CD3 for myeloperoxidase T-lymphoid, (MPO) for myeloid, expression of any two of CD14, CD64, CD11c, and lysozyme, or cytochemistry (esterase tests) for monocytic lineage.^[1] For B cell lineage assignment, the WHO has given two criteria, that is, either the strong expression of CD19 together with another B cell-associated marker or in cases with weak CD19, the expression of at least two B-lineage markers.^[1] In addition, the WHO recognizes two distinct categories,

This is an open access article distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 3.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as the author is credited and the new creations are licensed under the identical terms.

MPAL with the t(9;22)(q34;q11)/BCR-ABL1 and MPAL with t(v; 11q23)/mixed lineage leukemia (MLL) rearrangement. The remaining cases are designated as MPAL not otherwise specified. Although MPAL has been described in the literature, presentation as a solitary lesion without leukemic (bone marrow [BM]) involvement is rarely described. Identification of such cases using adequate immunophenotypic markers is very important as these cases had a poor prognosis, required intensive chemotherapy, and are resistant to treatment with a poor outcome.^[1] We are presenting two rare cases of mixed phenotypic blastic hematolymphoid neoplasms without leukemic involvement at disease presentation highlighting the importance of adequate immunophenotypic workup for the diagnosis of such lesions.

Case Reports

Case 1

An 8-year-old female presented with the right cheek swelling for 2 months with no additional symptoms. The swelling

How to cite this article: Ghodke K, Tembhare P, Patkar N, Subramanian PG, Arora B, Gujral S. A rare extramedullary and extralymphoid presentation of mixed phenotypic blastic hematolymphoid neoplasm: A study of two cases. Indian J Med Paediatr Oncol 2017;38:394-7.

Kiran Ghodke, Prashant Tembhare, Nikhil Patkar, PG Subramanian, Brijesh Arora¹, Sumeet Gujral

Hematopathology Laboratory, Department of Pathology, Tata Memorial Centre, ¹Department of Paediatric Oncology, Tata Memorial Centre, Mumbai, Maharashtra, India

Address for correspondence: Dr. Sumeet Gujral, Department of Pathology, Hematopathology Laboratory, 7th Floor, Annex Building, Tata Memorial Centre, Dr. E Borges Road, Parel, Mumbai - 400 012, Maharashtra, India. E-mail: s gujral@outlook.com



For reprints contact: reprints@medknow.com

was ill-defined, firm, and nontender. Her general and systemic examinations were normal. Positron emission tomography (PET) scan with F-18 fludeoxyglucose (FDG) revealed an FDG-avid soft tissue mass measuring $3.6 \text{ cm} \times 3.5 \text{ cm}$ with maximum standardized uptake value (SUV) of 11.88 in the right maxillary sinus with complete opacification and erosion of the adjacent superior alveolus with thinning of the walls of the maxillary sinus with an extramaxillary soft tissue component. Hematology values were as follows: hemoglobin (Hb) concentration - 13.1 g/dl; white blood cell (WBC) count - 7.8 \times 10⁹/L; platelets - 400 \times 10⁹/L; and peripheral blood smear (PBS) examination did not reveal any abnormal cells or blasts. Subsequent BM examination revealed normocellular to mildly hypocellular BM with trilineage hematopoiesis. No blasts or abnormal cells were identified. Cytology of cerebrospinal fluid also did not reveal any abnormal cells.

The right maxillary sinus biopsy showed diffuse proliferation of medium size cells with scant cytoplasm and blastic chromatin [Figure 1]. Mild pleomorphism in the size of the tumor cells with high mitotic activity was noted. On immunohistochemistry (IHC) [Figure 1], the tumor cells were positive for CD99 (Mic2), diffusely positive for B cell markers such as CD10, CD79a, PAX5 and also

expressed MPO and weakly CD45 (LCA) but negative for CD20, CD3, CD5, desmin, and chromogranin. In addition, nearly all tumor cells showed nuclear positivity for terminal deoxynucleotidyl transferase (TdT). Fluorescence *in situ* hybridization on interphase cells of formalin-fixed paraffin-embedded tissue revealed negative results for *BCR/ABL* using locus-specific identifier (LSI) *BCR/ABL* dual fusion translocation probe and for MLL using LSI MLL (11q23) break apart rearrangement probe; however, nearly 40% of cells revealed trisomy of chromosome 22. Thus, histological and IHC findings were consistent with bi/mixed phenotypic blastic hematolymphoid neoplasm with B-lymphoblastic lymphoma with coexpression of the myeloid lineage-specific marker and MPO.

She was started on treatment with institutional acute lymphoblastic leukemia protocol called MCP-841 with standard 4-drug induction followed by a consolidation block with intermediate-dose cytarabine known to be active in myeloid neoplasms, interim maintenance, and delayed intensification. After the end of induction, she had a good response in maxillary mass with complete metabolic and a partial morphological response with calcification of mass. She was continued on postinduction chemotherapy and received consolidative radiotherapy to the right maxilla in a dose of 25.2 grays in 14 fractions during interim

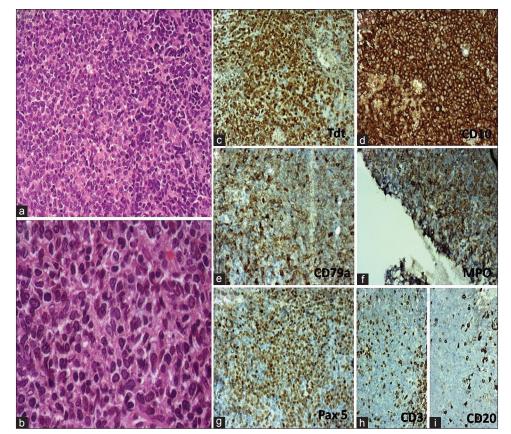


Figure 1: Case 1 - (a) Morphology showing diffuse proliferation of cells (H and E, ×400); (b) medium size cells with scant cytoplasm and blastic chromatin with mild pleomorphism in the size; (c) immunostain for terminal deoxynucleotidyl transferase showing nuclear expression (diaminobenzidine, ×400); (d-f) immunostain for CD10, CD79a, and myeloperoxidase showing membranous expression (diaminobenzidine, ×400); (g): immunostain for Pa × 5 showing nuclear expression (diaminobenzidine, ×400); (H and I): immunostain for CD3 and CD20 showing negative expression

maintenance. Her postradiation contrast-enhanced computed tomography scan revealed only thickening and sclerosis of the wall of maxillary sinus suggestive of healing. She is currently continuing on oral maintenance chemotherapy and is doing well after 3 maintenance cycles at 18-month follow-up.

Case 2

A 21-year-old male was being investigated for an acute onset of pain involving the left hip for 6-8 weeks. The magnetic resonance imaging revealed a permeative lesion involving the head and upper shaft of the femur. The needle core biopsy from the lesion was performed in another hospital which was reported as suspicious of a primitive neuroectodermal tumor and was referred to our institute. His laboratory workup in our hospital revealed normal complete blood count with Hb concentration - 14.2 g/dl; WBC count - 5.7×10^{9} /L; platelets - 278×10^{9} /L; and PBS did not reveal any abnormal cells or blasts. The serum lactate dehydrogenase was mildly raised to 216 U/L. PET scan with F-18 FDG revealed hypermetabolic disease involving the head, neck, and proximal one-third shaft of the left femur with maximum SUV of 7.6 with no obvious cortical destruction or soft tissue mass. In addition, the proximal end of the left fibula also showed an active disease with maximum SUV of 3.59. There was no evidence of active disease elsewhere in the body.

A biopsy of the lesion was reviewed which showed sheets of medium-sized tumor cells with scant cytoplasm and blastic chromatin with few crushing artifacts. Mild pleomorphism of tumor cells with focal apoptosis was noted. These features were consistent with a high-grade blastic neoplasm. On IHC, the tumor cells showed diffuse membranous expression of MIC2, whereas TdT showed weak, but distinct nuclear reactivity. In B-lymphoid lineage markers, CD10 and CD20 were diffusely positive while PAX5 was negative. In addition, blasts expressed MPO; however, CD117 did not reveal any positive expression.

The BM study was omitted initially as there was no evidence of any other sites or marrow involvement or uptake by PET scan. However, BM was done after a month of diagnosis, and it revealed 54% of blasts which were negative for cytochemical MPO. We performed eight-color flow cytometric immunophenotyping (FCI) using a comprehensive antibody panel [Table 1]. Cells were acquired on Navios (Beckman Coulter [BC]) flow cytometer using Navios software, and data were analyzed using Kaluza software (BC).

Immunophenotypic analysis [Figure 2] showed approximately 43% of viable cells were abnormal blasts. The abnormal blasts revealed a B-lymphoid lineage origin expressing CD10, CD19, CD20, and CD22 but negative for cytoplasmic CD79a. It also showed expression of CD34 and human leukocyte antigen–antigen D. In addition, they

Table 1: Comprehensive antibody panel	
B lymphoid markers	CD10, CD19, CD20, CD22,
	cytoplasmic CD79a
T lymphoid markers	CD2, sCD3, CD4, CD5, CD8,
	cytoplasmic CD3
Myeloid markers	CD11b, CD13, CD15, CD16, CD33,
-	CD117, cytoplasmic anti-MPO
Others	CD1a, CD34, CD38, CD45, CD56,
	CD58, CD123, HLADR

showed cytoplasmic MPO expression but were negative for rest of the other markers tested. BM trephine biopsy also showed a sheet of blasts expressing CD10, CD20, and MPO. Cytogenetics revealed no evidence of *BCR/ABL* fusion: t(9;22), AML1/ETO: t(8;21), and MLL rearrangement.

All features were consistent with bi/mixed phenotypic blastic hematolymphoid neoplasm as B-lymphoblastic lymphoma with coexpression of the myeloid marker and MPO that progressed to mixed phenotypic acute leukemia (B/myeloid leukemia) during a course of time.

Discussion

Most of acute leukemias are classified into myeloid, B-lymphoid, or T-lymphoid origin according to the antigenic expression of the blasts. However, in some cases, it is difficult to categorize them into specific lineage because of the expression of both lymphoid and myeloid lineage-specific antigens in the blast cells, and these leukemias have been designated as MPAL in the WHO.^[1] Although MPAL has been described in the literature, its presentation as a solitary lesion without leukemic involvement at the time of diagnosis is rarely described. To the best of our knowledge, only ten cases have been published in the literature,^[2-4] and ours is the first report of two cases from India. Largest case series of six cases of biphenotypic lymphoma (B and T) showed hypercellular BM with myeloid preponderance and eosinophilia with the t(8,13) association.^[2] One case had recurrent episodes of unexplained cervical lymphadenopathy over a period of several years and an acute onset of myasthenia gravis 1 year before patient's presentation with worsening respiratory difficulty,^[3] and another case had nontender neck lymph nodes for 9 months and was being evaluated for lymphoma.^[4] In these cases, tumor cells expressed both myeloid and T-lymphoid markers and lymph nodes being the most common site.

In the present study, both cases had extralymphatic bony involvement with no leukemic involvement. Notably, involvement in the second case, initially there was no leukemic involvement but progressed to leukemia during the course of the disease. Both cases had blasts showing unequivocal evidence of myeloid and B-lymphoid commitment as shown by the expression of lineage-specific antigens. Both cases could be diagnosed due to comprehensive immunophenotyping on biopsy. If we would

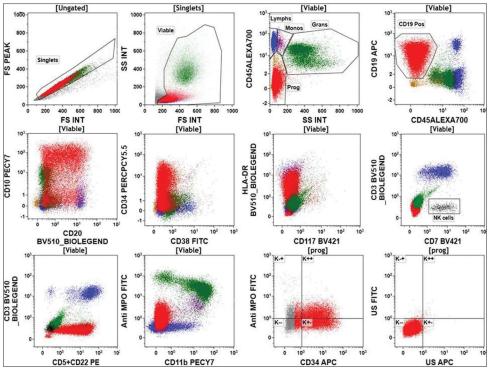


Figure 2: Immunophenotypic analysis of bone marrow specimen of Case 2. The dot plots show blasts (red dots) with positive expression for CD10, CD19, CD20, CD34, human leukocyte antigen–antigen D, and coexpression of CD22, and cytoplasmic myeloperoxidase. The blasts were negative for CD3, CD5, CD7, CD11b, CD38, CD45, and CD117

have been restricted our IHC to only lymphoid panel, these would have been diagnosed as B-lymphoblastic lymphoma. Both cases were difficult to categorize either into MPAL as the BM was not involved or into lymphoblastic lymphoma because of coexpression of myeloid differentiation. Therefore, we chose to classify them as a bi/mixed phenotypic blastic hematolymphoid neoplasm.

Conclusion

MPAL presenting as an extramedullary and extralymphatic lesion is extremely rare and difficult to categorize. Detailed immunophenotypic analysis on IHC or FCI is necessary for the diagnosis of such cases, and it is important to diagnose and classify them correctly as they harbor a poor prognosis.

Financial support and sponsorship

Nil.

Conflicts of interest

There are no conflicts of interest.

References

- Swerdlow S, Campo E, Harris N, Jaffe E, Pileri S, Stein H, *et al.* World Health Organization Classification of Tumours: Pathology and Genetics of Tumours of Haematopoietic and Lymphoid Tissues. Lyon, France: IARC Press; 2008. p. 150-5.
- Vega F, Medeiros LJ, Davuluri R, Cromwell CC, Alkan S, Abruzzo LV. t(8;13)-positive bilineal lymphomas: Report of 6 cases. Am J Surg Pathol 2008;32:14-20.
- Childs CC, Chrystal GS, Strauchen JA. Biphenotypic lymphoblastic lymphoma. An unusual tumor with lymphocytic and granulocytic differentiation. Cancer 1986;57:1019-23.
- Bayraktar UD, Barker J, Pereira D, Glick DZ, Byrne GE Jr., Escalón MP. An unusual presentation of extramedullary biphenotypic lymphoblastic lymphoma. Am J Hematol 2009;84:616-7.